

Clinical Pearls

Parvovirus B19-associated transient aplastic crisis following *P. falciparum* malaria and post-artesunate delayed haemolysis in a returning traveller

Ralph Huits^{ID}, MD PhD^{1,*}, Sabrina Van Ierssel, MD PhD², Anna Rosanas-Urgell, PhD³, Dorien Van den Bossche, MSc¹, and Emmanuel Bottieau, MD PhD¹

¹Department of Clinical Sciences, Institute of Tropical Medicine, Antwerp, Belgium, ²Department of General Internal Medicine, Antwerp University Hospital, Edegem, Belgium and ³Department of Biomedical Sciences, Institute of Tropical Medicine, Antwerp, Belgium

*To whom correspondence should be addressed. Email: rhuits@itg.be

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A 38-year-old, previously healthy Italian man was hospitalized at the University Hospital of Antwerp, Belgium, for recurrence of fever, 7 days after treatment for thick smear-positive *Plasmodium falciparum* malaria in Yaoundé, Cameroon (Day 0, Figure 1). He had not taken any malaria chemoprophylaxis. In Yaoundé, he did not have anaemia, but bilirubin levels were increased (8 mg/dL; normal value < 1.2 mg/dL). Parasitaemia levels were not documented. He had been treated with artesunate 2.4 mg/kg intravenously at 0, 12 and 24 h, followed by quinine 500 mg base orally every 8 h for 3 days, and he returned to Europe symptom-free. At the time of admission in Antwerp, the blood smear was negative for malaria. The patient was empirically treated with four tablets of dihydroartemisinin-piperaquine 320/40 mg once daily for 3 days. Fever (38.3°C) persisted for 1 week. On Day 14, the haemoglobin level dropped to 10.5 g/dL, with increasing LDH and undetectable haptoglobin. No schistocytes were detected in a peripheral blood smear. A direct antiglobulin test was positive for IgG, and negative for complement. Causes of anaemia considered at this stage included post-artesunate delayed haemolysis (PADH) and autoimmune haemolytic anaemia, possibly induced by malaria, underlying infections or antimalarial drugs.

Paradoxically, at a haematocrit value of 0.31 (normal range 0.39–0.49), the patient's reticulocyte count was 2.0 (absolute reticulocyte count 73 000 cells/ μ L) (Supplementary Figure S1). The reticulocyte production index (RPI), a correction of the reticulocyte count for red blood cell depletion and for reticulocyte

maturation time, was < 1, indicating that haemolysis was complicated by a hypoproliferative state.¹ From Day 17 through Day 20, mild neutropenia (1000–1500; normal 1800–7700 cells/ μ L) and presence of atypical lymphocytes were observed.

Serum tested negative for hepatitis B surface antigen, HCV, HIV and treponemal antibodies, and indicated past infections with CMV, EBV and dengue. On Day 15, IgM and IgG antibodies against parvovirus B19 (B19V) were detected. B19V-associated transient aplastic crisis (TAC) was suspected. B19V is a single-stranded DNA virus that spreads through respiratory secretions, blood-derived products or via materno–fetal transmission. Infection causes erythema infectiosum, hydrops fetalis, arthropathy and other pathologies, including TAC.² B19V targets erythroid progenitors and infection precipitates anaemia in haemolytic conditions (e.g. sickle cell disease). Infection confers lifelong protective immunity.²

When haemoglobin levels decreased to 6.1 g/dL on Day 19, two units of packed red blood cells were transfused. On Day 23, the RPI increased to 3.1, reflecting recovery from infection and increased compensatory erythropoiesis. After 3 months, anti-B19V IgG persisted and IgM antibodies were no longer detected.

Falciparum malaria and B19V co-infections have been identified as important contributors to anaemia in endemic countries,³ but are rarely reported in travellers.⁴ B19V prevalence in febrile returning travellers may be underreported, because B19V diagnostics are not part of the routine work-up. PADH in returned travellers rarely leads to transfusion-dependent anaemia.⁵





Day	0		3			7		10		14		17		21		// 4W //	9W //	15W
Fever																		
Malaria*	+					-		-			-							
Treatment	A	Q	Q	Q		DP	DP	DP										
Atyp. lympho.																		
B19V IgG								+			+							+
B19V IgM								+/-			+							-
Blood transfusion																		
Ht (L/L)						0.32		0.35		0.31	0.31	0.24	0.19	0.19	0.22	0.21		0.40
Hb (g/dL)	14.0					11.6		12.3		10.5		7.9		6.1	7.5	7.0	9.9	14.0
Haptoglobin (g/L)								1.18		U		U						
Bilirubin (mg/dL)	8.0							2.0			2.8	2.6		3.0				
Retic. Count (%)								4.2		2.0	2.6	3.8	5.3		7.1	12.1		
RPI (%)								2.2		0.9	1.2	1.0	0.9		1.8	2.8		

Figure 1. Clinical and laboratory data of a traveller with B19V-associated TAC following *P. falciparum* malaria and post-artesunate delayed haemolysis. Fig. 1. (Legend) Day 0 is the time of malaria diagnosis (Cameroon, 29 June 2021). W denotes weeks. Malaria* denotes 'thick blood smear'; Treatment (for malaria): A, artesunate; Q, quinine; DP, dihydroartemisinin-piperaquine; Atyp. lympho., atypical lymphocytes detected; B19V, parvovirus B19; IgG, immunoglobulin G; IgM, immunoglobulin M; Ht, haematocrit; Hb, haemoglobin; ▼ Blood transfusion of two units of red blood cells; retic. Count: reticulocytes, expressed as a percentage of red blood cells; + positive; - negative; +/- equivocal result; U, undetectable; RPI, reticulocyte production index. The formula used to calculate the RPI is: $RPI = (\text{reticulocyte count} \times Ht/0.45)/\text{maturation correction}$, where 0.45 is the normal Ht-value; the maturation correction reflects the longer life span of premature reticulocytes and is corrected for Ht-values ≥ 0.40 (L/L) by factor 1.0, for Ht-values 0.30–0.39 by 1.5, for Ht-values 0.20–0.29 by 2.0 and for Ht-values < 0.20 by 2.5. Interpretation: an RPI $> 3\%$ shows a normal marrow response to anaemia. An RPI $< 2\%$ is an inadequate response to correct anaemia¹

However, a B19V-associated TAC that impairs erythropoiesis for 7 to 10 days after malaria-related haemolysis or PADH is potentially life-threatening. TAC can be adequately managed by timely blood transfusions.² Reticulocytopenia and an RPI $< 2\%$ following *P. falciparum* malaria should alert clinicians to concomitant B19V infection.

Supplementary data

Supplementary data are available at *JTM* online.

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Author roles

R.H. conceptualized and developed the initial draft manuscript. All authors reviewed and approved the final manuscript.

References

1. Hillman RS. Characteristics of marrow production and reticulocyte maturation in normal man in response to anemia. *J Clin Invest* 1969; **48**:443–53.
2. Qiu J, Söderlund-Venermo M, Young NS. Human parvoviruses. *Clin Microbiol Rev* 2017; **30**:43–113.
3. Pasvol G. Parvovirus infection, malaria, and anemia in the tropics – a new hidden enemy? *J Infect Dis* 2006; **194**:141–2.
4. Scarlata F, Gianelli E, Miceli S *et al*. Acute parvovirus B19 infection and anemia during plasmodium falciparum malaria. *Clin Infect Dis* 2002; **35**:1449–51.
5. Jauréguiberry S, Thellier M, Ndour PA *et al*. Delayed-onset hemolytic anemia in patients with travel-associated severe malaria treated with Artesunate, France, 2011–2013. *Emerg Infect Dis* 2015; **21**: 804–12.